

FAECAL STEROID ANALYSIS IN FREE-RANGING WHITE RHINOCEROSSES (*Ceratotherium simum*) ON A GAME RANCH IN NAMIBIA

**B. MRAZ¹, C. WALZER², P. REINHARDT³, J. MATTHAEI³, M. JAGO⁴ and
F. SCHWARZENBERGER¹**

Affiliation:

1. Dept of Natural Sciences, Inst. Biochemistry, Univ. Vet. Med., A-1210-Vienna, Austria
2. Salzburg Zoo Hellbrunn, Salzburg, Austria
3. Otjiwa Game Ranch, Otjiwarongo, Namibia
4. Otjiwarongo Vet. Clinic, Otjiwarongo, Namibia

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Extended abstract

The reproductive problems of captive male and female white rhinoceroses were summarised at recent wildlife conferences (2, 4, 5). Approximately 60% of female captive white rhinoceroses are a-cyclic (6). The reasons are 1) females with no luteal activity (true flatliners) and 2) females with persistent luteal activity. Females from the last group are nulli-parous, usually over 12 years of age and their uterus is affected by severe pathologies (cysts) resulting in infertility. In contrast, cysts are usually not present in primi- or multi-parous females (2, 4). Male infertility in captive white rhinoceroses is a severe additional problem and cause for the low reproductive rate. Our results imply that more than 2 males in the same institution can suppress each other's fertility (1).

The objectives of this study were to determine whether reproductive problems are present in wild populations. Study site was Otjiwa game ranch (size: 100 km²), Namibia. The number of rhinoceroses on this farm varied between 21 and 26 animals during the past 3 years. The herd consists of about 8 breeding females with their offspring, and 1 – 2 bulls of breeding age. Until recently only one dominant bull was kept on the farm. Severe fighting between this dominant bull and the younger bulls made removal of bulls over the age of 4-5 y necessary. Currently 2 bulls of breeding age are kept at the farm and sample collection from these bulls is in progress in order to elucidate to what extent their territorial status is reflected in faecal androgen concentrations.

All females of breeding age on Otjiwa game ranch are usually with calves and age at first calving was around 7.5 years. Calving intervals of about 2.5 years indicate that infertility conditions, comparable to captive white rhinoceroses in the EEP or SSP, are not present. In order to monitor oestrous cyclicity in this semi-wild population, fresh dung samples from the females were serially collected after tracking by game scouts over the past 3 years. Samples were stored in methanol and lyophilised prior to analysis with an established 20-oxo-pregnane assay (6). Average individual sample collection interval was about 20 d (range 1-100 d). This long interval and the fact that females seem to become pregnant after a short period of lactational an-oestrus hindered statements on cyclicity. However, pregnancy diagnosis, even from single samples was 100% accurate after Day 120.

The major problem in our study is the identification of collected samples, as farm size and brush coverage made tracking difficult. Profiles of faecal progesterone metabolite concentrations from

pregnant animals indicate that the game scouts misidentified about 20-25 % of samples. As ear notches and radio transmitters are currently not available with our study animals, a digital spoor identification technique is presently under investigation (3). This non-invasive technique was recently developed for identifying individual black rhino from their footprints (spoor) and is being adapted for the identification of the white rhinoceroses on Otjiwa game ranch. For spoor identification, digital images were taken of the hind spoor from tracks, and landmark points were manually placed on the spoor image. From these digital images, measurements (lengths and angles) were generated using customised software (3). The limitation of this technique is the difficulty of the practical applications by the African game scouts, who have no experience in using computers. In addition, the quality of the spoor images varies with dry or rainy season. In summary, use of faecal steroid analysis in a semi-wild white rhinoceros population is hampered by difficulties in the identification of collected samples. Therefore we are currently investigating whether sample identification can be improved by applying a digital spoor identification technique.

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